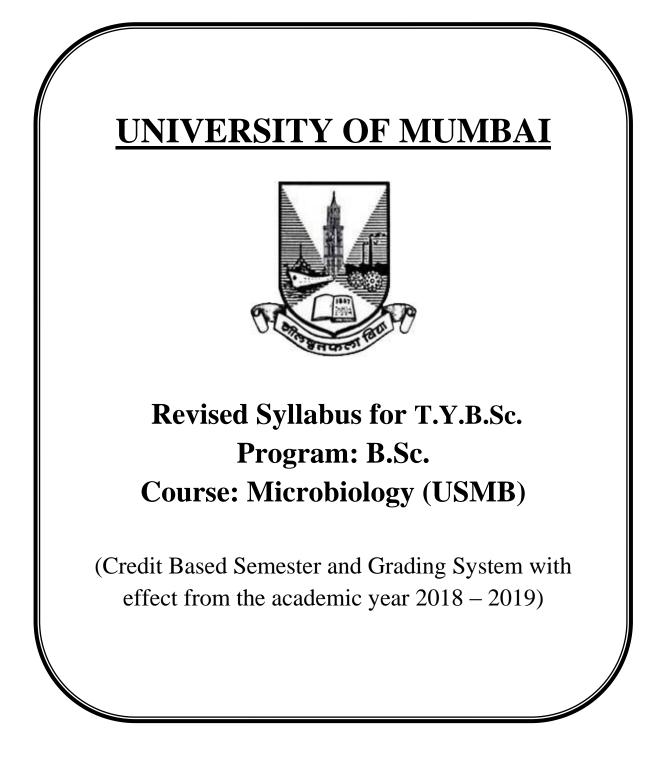
Item No.



PREAMBLE

The Choice Based Credit system was introduced by Mumbai University from 2016 - 2017. The process was initiated by restructuring the F.Y.B.Sc. syllabus and the paper pattern according to the CBCS pattern and its implementation in the same year i.e. 2016 - 17.

This was followed by revision of S.Y.B.Sc. syllabus and paper pattern in the year 2017 - 2018.

The revised S.Y.B.Sc. syllabus gave an opportunity to the Microbiology students to opt for Paper III of any subject other than Microbiology. Likewise S.Y.B.Sc. students of other subjects could opt for Microbiology Paper III. This gave them the option to choose from diversity of applied sciences.

In continuation with this, the T.Y.B.Sc. syllabus is being revised in the year 2018 - 2019. The existing paper pattern will also be accordingly revised.

Keeping in tune with the revised syllabus, the committee has ensured that there is a continuous flow of information and latest advances in the subject imparted to the students. Hence some of the modules of the earlier syllabus have been upgraded, while some new modules have been added to the syllabus in order to bridge the knowledge gap of the learner from S.Y.B.Sc. to T.Y.B.Sc.

The syllabus is aimed at equipping the students with basic knowledge in various branches of Microbiology such as Microbial Genetics, Molecular Biology, Virology, Medical Microbiology, Immunology, Microbial Biochemistry and Industrial Microbiology. Additionally, it also makes students aware of interdisciplinary sciences such as Bioinformatics and Bioinstrumentation.

In all, the students offering Microbiology as a single major subject that is Six units pattern, will study eight courses of theory and practicals compulsory during Semester V and Semester VI together, while students opting for double major subject that is Three units pattern, will have four courses of theory and practicals compulsory during Semester V and Semester VI together.

The courses for six units will comprise of the following:

- 1) USMB 501 and USMB 601
- 2) USMB 502 and USMB 602
- 3) USMB 503 and USMB 603
- 4) USMB 504 and USMB 604

The courses for three units will comprise of the following:

- 1) USMB 501 and USMB 601
- 2) USMB 502 and USMB 602

The approach towards designing this syllabus has been to retain the classic concepts of Microbiology as well as keeping abreast with the latest discoveries in Microbiology and other interdisciplinary fields.

In conclusion, the revised syllabus aims at inculcating a spirit of learning and kindling curiosity towards the subject in the minds of learners, resulting in their pursuit of higher education in Microbiology.

T.Y.B.Sc. MICROBIOLOGY THEORY

COURSE CODE	TITLE	CREDITS AND LECTURES / SEM		
USMB501	Microbial Genetics	2.5 Credits (60 Lectures)		
Unit I	DNA Replication	15 Lectures		
Unit II	Transcription, Genetic Code & Translation	15 Lectures		
Unit III	Mutation and Repair	15 Lectures		
Unit IV	Genetic Exchange & Homologous Recombination	15 Lectures		
USMB502	USMB502 Medical Microbiology & Immunology: Part - I			
Unit I	Bacterial Strategies for Evasion and Study of a Few Diseases	15 Lectures		
Unit II	Study of a Few Diseases with Emphasis on Cultural Characteristics of the Etiological agent, Pathogenesis, Laboratory Diagnosis and Prevention.	15 Lectures		
Unit III	General Immunology - I	15 Lectures		
Unit IV	General Immunology - II	15 Lectures		
USMB503	Microbial Biochemistry: Part - I	2.5 Credits (60 Lectures)		
Unit I	Biological Membranes & Transport	15 Lectures		
Unit II	Bioenergetics & Bioluminescence	15 Lectures		
Unit III	Methods of Studying Metabolism & Catabolism of Carbohydrates	15 Lectures		
Unit IV	Fermentative Pathway & Anabolism of Carbohydrates	15 Lectures		

(SEMESTER V)

USMB504	Bioprocess Technology: Part - I	2.5 Credits (60 Lectures)				
Unit I	Upstream Processing - I	15 Lectures				
Unit II	Upstream Processing - II	15 Lectures				
Unit III	Fermentation Modes, Equipments and Instruments	15 Lectures				
Unit IV	Traditional Industrial Fermentations	15 Lectures				

N.B.

- I. Each theory period shall be of 48 minutes duration. Theory component shall have 240 instructional periods plus 240 notional periods per semester which is equal to 384 learning hours. For theory component the value of One Credit is equal to 38.40 learning hours.
- II. Each practical period shall be of 48 minutes duration. Practical component shall have 240 instructional periods plus 60 notional periods per semester which is equal to 240 learning hours. For practical component the value of One Credit is equal to 40 learning hours.

T.Y.B.SC. MICROBIOLOGY THEORY (SEMESTER V)

MICROBIAL GENETICS (USMB-501)

LEARNING OBJECTIVES

Microbial Genetics (USMB-501) is a course in Genetics for T.Y.B.Sc. undergraduate students in Semester V that deals with various concepts of Genetics.

The learning objectives include the following:

- 1. **DNA Replication:** The learner will understand the events occurring in both Prokaryotic and Eukaryotic DNA replication, with a focus on the involvement of Proteins and Enzymes at the cellular level. The topic will also include the assembly of Eukaryotic chromosome.
- 2. **Transcription, Genetic Code and Translation:** This module aims at the learner understanding the basis of gene expression and the Central Dogma and the molecular basis of protein synthesis in Prokaryotes and Eukaryotes. The module deals with the structure and properties of different forms of RNA, maturation of RNA and RNA splicing.
- 3. **Mutation and DNA repair:** The molecular basis and types of mutation, their cause, effect and DNA repair is studied. The basic concepts related to molecular biology are explained.
- 4. **Genetic exchange:** This module includes the study of various mechanisms of gene transfer in bacteria. It also provides insight into the mechanisms of genetic recombination. The module deals with the Genetics of bacteria and bacteriophages, development of new strains and genetic mapping.
- 5. Practicals

The laboratory techniques and experiments based on these topics will give students hands on competence in fundamental molecular biology experiments.

LEARNING OUTCOMES:

- **DNA Replication**: The learner will understand the sequence of events, mechanism, enzymes and proteins involved in replication of DNA in prokaryotes and eukaryotes.
- **Transcription, Genetic Code and Translation:** The student will know the central dogma of biology its two-step transcription and translation, maturation of RNA.
- **Mutation and DNA repair**: The learner will know the concept of mutation, its types, causes and their effects. This module will also make them understand types of mutagens, damage to DNA due to mutagenesis, various mechanisms of DNA repair.
- **Genetic exchange**: The student shall understand the various mechanisms of gene transfer in bacteria and genetic recombination.
- **Practicals**: The students will acquire skill to perform the laboratory techniques and experiments based on the above topics.

MICROBIAL GENETICS (USMB-501): DETAIL SYLLABUS

	Title	Lectures / Semester	Notional Periods
	Unit I: DNA Replication	15 L	15
1.1.	Historical perspective - Conservative, dispersive, semi-conservative, bidirectional and semi-discontinuous, Theta model of replication.	3 L	
1.2.	Prokaryotic DNA replication - Details of molecular mechanisms involved in Initiation, Elongation and Termination	4 L	
1.3.	Enzymes and proteins associated with DNA replication - Primase, Helicase, Topoisomerase, SSB, DNA polymerases, Ligases, Ter and Tus proteins.	3 L	
1.4.	Eukaryotic DNA replication - Molecular details of DNA synthesis, replicating the ends of the chromosomes assembling newly replicated DNA into nulcleosomes.	4 L	
1.5.	Rolling circle mode of DNA replication	1 L	
	Unit II: Transcription, Genetic Code and Translation	15 L	15
2.1	Central Dogma: An Overview, Transcription process, Transcription in bacteria - Initiation of transcription at promoters, elongation of an RNA chain, termination of an RNA chain	3 L	
2.2	Transcription in Eukaryotes - Eukaryotic RNA polymerase, Transcription of protein- coding genes by RNA polymerase II, Transcription initiation, The structure and production of Eukaryotic mRNAs, Production of mature mRNA in Eukaryotes, Processing of Pre-mRNA to mature mRNA. Self Splicing of Introns, RNA editing	5 L	
2.3	Genetic code - Nature of genetic code and characteristics of genetic code.	2 L	
2.4	Translation process - Transfer RNA, structure of tRNA, tRNA genes, Recognition of the tRNA anticodon by the mRNA codon, Adding of amino acid to tRNA, Ribosomal RNA and Ribosomes, Ribosomal RNA Genes, Initiation of translation, Initiation in Bacteria, Initiation in eukaryotes, Elongation of the polypeptide chain, termination of translation, protein sorting in the cell.	5 L	
	Unit III: Transcription, Genetic Code and Translation	15 L	15
3.1	 Mutation 3.1.1 Terminology: alleles, homozygous, heterozygous, genotype, phenotype, Somatic mutation, Germline mutation, Gene mutation, Chromosome mutation, phenotypic lag, hotspots and mutator genes 	1 L	

	3.1.2	Fluctuation test.	1 L	
	3.1.3	Types of mutations: Point mutation, reverse mutation, suppressor mutation, frameshift mutation, conditional lethal mutation, base pair substitution, transition, transversion, missense mutation, nonsense mutation, silent mutation, neutral mutation, pleiotropic mutations.	3 L	
	3.1.4	Causes of mutation: Natural/spontaneous mutation replication error, depurination, deamination. Induced mutation: principle and mechanism with illustrative diagrams for:	4 L	
		3.1.4.1 Chemical mutagens - base analogues, nitrous acid, hydroxyl amine, intercalating agents and alkylating agents.		
		3.1.4.2 Physical mutagen		
		3.1.4.3 Biological mutagen (only examples)		
	3.1.5	Ames test	1 L	
	3.1.6	Detection of mutants	1 L	
3.2	DNA R	enair	4 T	
J. 4	3.2.1	Mismatch repair,	4 L	
	3.2.2	Light repair		
	3.2.2	0 1		
	3.2.4			
	3.2.5 Nucleotide excision repair			
	3.2.6	SOS repair		
	Unit I	V: Genetic Exchange & Homologous Recombination	15 L	15
4.1	Genetic	e analysis of Bacteria	1 L	
4.2	C 4-	ansfer mechanisms in bacteria		
4.2	4.2.1	Transformation	3 L	
	4.2.1	4.2.1.1 Introduction and History	31	
		4.2.1.1 Introduction and History 4.2.1.2 Types of transformation in prokaryotesNatural		
		transformation in <i>Streptococcus pneumoniae</i> ,		
		Haemophilus influenzae, and Bacillus subtilis.		
		4.2.1.3 Mapping of bacterial genes using transformation.		
		4.2.1.4 Problems based on transformation.		
	4.2.2	Conjugation	5 L	
	1.4.4	4.2.2.1 Discovery of conjugation in bacteria	ЭL	
		4.2.2.2 Properties of F plasmid/Sex factor		
		4.2.2.3 The conjugation machinery		
		4.2.2.4 Hfr strains, their formation and mechanism of		
		conjugation		
		4.2.2.5 F' factor, origin and behavior of F' strains,		

		4.2.2.6	Sexduction. Mapping of bacterial genes using conjugation		
			(Wolman and Jacob experiment).		
		4.2.2.7	Problems based on conjugation		
	4.2.3	Transduc	ction		
		4.2.3.1	Introduction and discovery	3 L	
		4.2.3.2	Generalized transduction	012	
		4.2.3.3	Use of Generalized transduction for mapping genes		
		4.2.3.4	Specialized transduction		
		4.2.3.5	Problems based on transduction		
4.3	Recom	bination i	n bacteria	3 L	
	4.3.1	General/	Homologous recombination		
	4.3.2	Molecula	ar basis of recombination		
	4.3.3	Holliday	model of recombination (Single strand DNA break		
		model or	ıly)		
	4.3.4				
	4.3.5	Site –spe	cific recombination		

MEDICAL MICROBIOLOGY & IMMUNOLOGY: PART-I

TEXT BOOKS AND REFERENCE BOOKS (SEMESTER V)

Course Code: USMB501

Text books:

- 1. Peter J. Russell (2006), "I Genetics-A molecular approach", 2nd edition.
- 2. Benjamin A. Pierce (2008), "Genetics a conceptual approach", 3rd edition, W. H. Freeman and company.
- 3. R. H. Tamarin, (2004), "Principles of genetics", Tata McGraw Hill.
- 4. D. Nelson and M. Cox, (2005), "Lehninger's Principles of biochemistry", 4th edition, Macmillan worth Publishers.
- 5. M. Madigan, J. Martinko, J. Parkar, (2009), "Brock Biology of microorganisms", 12th edition, Pearson Education International.
- 6. Fairbanks and Anderson, (1999), "Genetics", Wadsworth Publishing Company.
- 7. Prescott, Harley and Klein, "Microbiology", 7th edition Mc Graw Hill international edition.
- 8. Robert Weaver, "Molecular biology", 3rd edition. Mc Graw Hill international edition.
- 9. Nancy Trun and Janine Trempy, (2004), "Fundamental bacterial genetics", Blackwell Publishing
- 10. Snustad, Simmons, "Principles of genetics", 3rd edition. John Wiley & sons, Inc.

Reference books:

- 1. Benjamin Lewin, "Genes IX", Jones and Bartlett publishers.
- 2. JD Watson, "Molecular biology of the gene", 5th edition.

T.Y.B.Sc. MICROBIOLOGY THEORY

(SEMESTER VI)

COURSE CODE	TITLE	CREDITS AND LECTURES / SEM
USMB601	rDNA Technology, Bioinformatics & Virology	2.5 Credits (60 Lectures)
Unit I	Recombinant DNA Technology	15 Lectures
Unit II	Applications of rDNA Technology & Bioinformatics	15 Lectures
Unit III	Regulation & Basic Virology	15 Lectures
Unit IV	Advanced Virology	15 Lectures
USMB602	Medical Microbiology & Immunology: Part - II	2.5 Credits (60 Lectures)
Unit I	Study of a Few Diseases with Emphasis on Cultural Characteristics of the Etiological Agent, Pathogenesis, Laboratory Diagnosis and Prevention.	15 Lectures
Unit II	Chemotherapy of Infectious Agents	15 Lectures
Unit III	Immunology - I	15 Lectures
Unit IV	Immunology – II	15 Lectures
USMB603	Microbial Biochemistry: Part - II	2.5 Credits (60 Lectures)
Unit I	Lipid Metabolism & Catabolism of Hydrocarbons	15 Lectures
Unit II	Metabolism of Proteins and Nucleic Acids.	15 Lectures
Unit III	Metabolic Regulation	15 Lectures
Unit IV	Prokaryotic Photosynthesis & Inorganic Metabolism	15 Lectures
USMB604 Bioprocess Technology: Part - II		2.5 Credits (60 Lectures)
Unit I	Downstream Processing	15 Lectures
Unit II	Advances in Bioprocess Technology	15 Lectures
Unit III	Quality Assurance, Quality Control, Instrumentation and Bioassay	15 Lectures
Unit IV	Industrial Fermentations	15 Lectures

T.Y.B.SC. MICROBIOLOGY THEORY (SEMESTER V) rDNA TECHNOLOGY, BIOINFORMATICS & VIROLOGY (USMB-601)

LEARNING OBJECTIVES

rDNA technology, Bioinformatics and Virology, USMB 601 is a course for T.Y.B.Sc. in Semester VI Microbiology students which deal with the following:

- 1. **The rDNA technology:** This module deals with the basic steps in gene cloning, vectors, model organisms, methods of transformation and screening and identification of recombinant cells.
- 2. **Application of rDNA technology and Bioinformatics:** This module will empower the student to understand the basic techniques in Recombinant DNA technology along with their applications. Bioinformatics is the basic tool in understanding Cells at the genomic and proteomic levels. Inclusion of Bioinformatics in this module will empower the learner with insilico analytical techniques.
- 3. Gene Regulation and Basic Virology: This module will make the students understand the genetic basis of regulation and operon control through the involvement of regulatory proteins. The study of Basic Virology will emphasise on the structure, classification and general modes of replication of viruses.
- 4. Advanced Virology: This module deals with basic structure and life cycle of different viruses and cultivation of viruses. It also comprises of basic study on Prions, Viriods and viruses causing cancer.

LEARNING OUTCOMES:

- **r DNA technology:** This module will make the student understand the methods to construct recombinant DNA molecules, also know the tools required like vectors, restriction enzymes etc.
- Application of rDNA technology and Bioinformatics: The learner will know about applications of r DNA technology, through bioinformatics the student will understand the use of databases and software tools for understanding biological data.
- Gene Regulation and Basic Virology: The student will know about gene expression in prokaryotes, operon as a unit of gene regulation, regulation of gene expression in prokaryotes and bacteriophages. The student will also understand about general structure, life cycle and classification of viruses.
- Advanced Virology: The learner will understand the basic structure and life cycle of different viruses and their cultivation. The student will get basic knowledge on Prions, Viriods and viruses causing cancer.
- **Practicals**: The students will acquire skill to perform the laboratory techniques and experiments based on the above topics. The students will understand computational biology and insilico analytical techniques.

rDNA TECHNOLOGY, BIOINFORMATICS & VIROLOGY

(USMB-601): DETAIL SYLLABUS

	Title	Lectures / Semester	Notional Periods
	Unit I: Recombinant DNA Technology	15 L	15
1.1	Branches of Genetics	1 L	
	1.1.1 Transmission genetics		
	1.1.2 Molecular genetics		
	1.1.3 Population genetics		
	1.1.4 Quantitative genetics		
1.2	Model Organisms	2 L	
	1.2.1 Characteristics of a model organism	2 L	
	1.2.2 Examples of model organisms used in study		
	1.2.3 Examples of studies undertaken using prokaryotic and	1	
	eukaryotic model organisms		
1.3	Plasmids		
	1.3.1 Physical nature	2 L	
	1.3.2 Detection and isolation of plasmids		
	1.3.3 Plasmid incompatibility and Plasmid curing		
	1.3.4 Cell to cell transfer of plasmids		
	1.3.5 Types of plasmids		
	1.3.6 Resistance Plasmids, Plasmids encoding Toxins and other	r	
	Virulence characteristics, Colfactor, Degradative plasmids		
1.4	Transposable Elements in Prokaryotes	2 L	
1.7	1.4.1 Insertion sequences		
	1.4.2 Transposons: Types, Structure and properties, Mechanism of	f	
	transposition, Integrons		
1.5	Basic steps in Gene Cloning.	1 L	
1.6	Cutting and joining DNA molecules - Restriction and modification systems, restriction endonucleases, DNA ligases	a 3 L	
1.7	Vectors	3 L	
	1.7.1 Plasmids as cloning vectors. plasmid vectors, pBR322 vector	JL	
	1.7.2 Cloning genes into pBR322		
	1.7.3 Phage as cloning vectors, cloning genes into phage vector		
	1.7.4 Cosmids		
	1.7.5 Shuttle vectors		
	1.7.6 YAC		
	1.7.7 BAC		
1.8	Methods of transformation	1 L	

	Unit II: Applications of rDNA Technology & Bioinformatics	15 L	15
2.1	PCR - basic PCR and different types of PCR (Reverse transcriptase PCR, Real time quantitative PCR)	2 L	
2.2	Basic techniques2.2.1Southern, Northern and Western blotting.2.2.2Autoradiography (explain the term	2 L	
2.3	Screening and selection methods for identification and isolation of recombinant cells	2 L	
2.4	Applications of recombinant DNA technology: Site specific mutagenesis of DNA, Uses of DNA polymorphism, STRS and VNTRS, DNA molecular testing for human genetic diseases (Only RFLP), DNA typing, gene therapy, Genetic engineering of plants and animals.	4 L	
2.5	 Bioinformatics 2.5.1 Introduction 2.5.2 Definition, aims, tasks and applications of Bioinformatics. 2.5.3 Database, tools and their uses – 2.5.3.1 Importance, Types and classification of databases 2.5.3.2 Nucleic acid sequence databases- EMBL, DDBJ, GenBank, GSDB, Ensembl and specialized Genomic resources. 2.5.3.3 Protein sequence databases-PIR, SWISS-PROT, TrEMBL NRL-3D.Protein structure databases-SCOP, CATH, PROSITE, PRINTS and BLOCKS. KEGG. 2.5.4 Explain the terms: Transcriptome, Metabolomics, Pharmacogenomics, Phylogenetic analysis, Phylogenetic tree, Annotation, Genomics- structural and functional and comparative genomics, Proteomics - structural and functional proteomics, Sequence alignment - global v/s local alignment, 	5 L	
	FASTA, BLAST (Different types of BLAST) Unit III: Regulation & Basic Virology	15 L	15
3.1	A) Lac operon and problems on Lac operonB) Trp operon	13 L 7 L	13
3.2	Regulation of lytic and lysogenic pathway of lambda phage	3 L	
3.3	Viral architecture - Capsid, viral genome and envelope	2 L	
3.4	Viral classification (Baltimore classification)	1 L	
3.5	Viral replication cycle - Attachment, penetration, uncoating, types of viral genome, their replication, assembly, maturation & release.	2 L	

		1	Unit IV: Advanced Virology	15 L	15	
4.1			V, T4, Influenza virus, HIV. Life cycle of T4 phage, Virus and HIV in detail.	5 L		
4.2	labora	vation of vation of vation of vation of vatication of the second	3 L			
4.3	Visua	lization an	d enumeration of virus particles	3 L		
	4.3.1		nent of infectious units	υĽ		
		4.3.1.1	Plaque assay			
		4.3.1.2	Fluorescent focus assay			
		4.3.1.3	Infectious center assay			
		4.3.1.4	Transformation assay			
		4.3.1.5	Endpoint dilution assay.			
	4.3.2		nent of virus particles and their components			
		4.3.2.1	Electron microscopy			
		4.3.2.2	Atomic force microscopy			
		4.3.2.3	Haemagglutination			
		4.3.2.4	Measurement of viral enzyme activity.			
4.4	cance	of viruses r cell, Huma itis B and C	2 L			
4.5		Prions: Defination, Examples of diseases caused by prions, Kuru, PrP protein and protein only hypothesis				
4.6	Viroi	ds		1 L		

TEXT BOOKS AND REFERENCE BOOKS (SEMESTER VI)

Course Code: USMB601

Text books:

- 1. Peter J. Russell (2006), "I Genetics-A molecular approach", 2nd edition.
- 2. Benjamin A. Pierce (2008), "Genetics a conceptual approach", 3rd edition, W. H. Freeman and company.
- 3. R. H. Tamarin, (2004), "Principles of genetics", Tata McGraw Hill.
- 4. M. Madigan, J. Martinko, J. Parkar, (2009), "Brock Biology of microorganisms", 12th edition, Pearson Education International.
- 5. Fairbanks and Anderson, (1999), "Genetics", Wadsworth Publishing Company.
- 6. Prescott, Harley and Klein, "Microbiology", 7th edition Mc Graw Hill international edition.
- Edward Wagner and Martinez Hewlett, (2005) "Basic Virology", 2nd edition, Blackwell Publishing
- 8. Teri Shors,.(2009), "Understanding viruses", Jones and Bartlett publishers.
- 9. S.Ignacimuthu, (2005), "Basic Bioinformatics", Narosa publishing house.
- 10. Robert Weaver, (2008), "Molecular biology", 3rd edition, Mc Graw Hill international edition.
- 11. Primrose and Twyman, (2001), "Principles of gene manipulation and genomics", 6th edition, Blackwell Publishing
- 12. Arthur Lesk, (2009), "Introduction to Bioinformatics", 3rd edition, Oxford University Press
- 13. Snustad, Simmons, "Principles of genetics", 3rd edition. John Wiley & sons, Inc.
- 14. A textbook of biotechnology R. C. Dubey 4th edition. S. Chand.

Reference books:

- 1. Flint, Enquist, Racanillo and Skalka, "Principles of virology", 2nd edition. ASM press.
- 2. T. K. Attwood & D. J. Parry-Smith, (2003), "Introduction to bioinformatics", Pearson education
- 3. Benjamin Lewin, (9th edition), "Genes IX", Jones and Bartlett publishers.
- 4. JD Watson, "Molecular biology of the gene", 5th edition.

Modality of Assessment Assessment pattern for theory

Scheme of Examination

The learner's Performance shall be assessed by conducting the Semester End Examinations with 100% marks

Semester End Theory Assessment - 100%

100 marks

- 1. Duration These examinations shall be of **3 hours** duration.
- 2. Theory question paper pattern :
 - i. There shall be **five questions** each of **20** marks (with internal options)
 - ii. Question one will be based on unit one, question two on unit two, question three on unit three and question four on unit four. Question five will have questions from all four units of the syllabus.
 - iii. Each of the main questions one to four will be subdivided into two sub-questions "A" and "B". Sub-question "A" will have four questions (of 6 marks each) out of which any two will be attempted. Total marks allotted to sub-question "A" will be 12 marks. Sub-question "B" will be 'Do as directed (attempt eight out of twelve)'. Each question in Sub-question "B" will be of one mark each. Total marks allotted to "B" sub-question will be 8 marks. Main question five will have six questions (of 5 marks each) out of which any four will be attempted, total 20 marks.
 - iv. All questions shall be **compulsory** with internal choice within the questions.
 - v. The allocation of marks will depend on the weightage of the topic.

Passing Standard:

The learners to pass a course shall have to obtain a minimum of 40% marks in aggregate for each course and 40% marks in **Semester End Examination (i.e. 40 out of 100) separately**, to pass the course and **minimum of Grade E** in each project, wherever applicable, to pass a particular semester.

Practical Examination Pattern:

External (Semester end practical examination):-

Sr.No.	Particulars/ paper	Marks
1.	Laboratory work	40
2.	Journal	05
3.	Viva	05

Semester V:

The students are required to present a duly certified journal for appearing at the practical examination, failing which they will not be allowed to appear for the examination.

In case of loss of Journal and / or Report, a Lost Certificate should be obtained from the Head of the Department / Co-ordinator of the department; failing which the student will not be allowed to appear for the practical examination.

Semester VI

The students are required to present a duly certified journal for appearing at the practical examination, failing which they will not be allowed to appear for the examination.

In case of loss of Journal and/ or Report, a Lost Certificate should be obtained from the Head of the Department/ Co-ordinator of the department; failing which the student will not be allowed to appear for the practical examination.

Overall Examination and Marks Distribution Pattern

Course code	Practical Syllabus	Credits & lectures
USMBP05	Based on USMB501 and USMB502 of Semester V	Credits 3 (8 periods/week) = 120 periods/semester
USMBP06	Based on USMB503 and USMB504 of Semester V	Credits 3 (8 periods/week) = 120 periods/semester

Semester V

Course	USMB- 501	USMB- 502	USMB- 503	USMB- 504	Grand Total
Theory	100	100	100	100	400
Practicals	50	50	50	50	200

Semester VI

Course	USMB- 601	USMB- 602	USMB- 603	USMB- 604	Grand Total	
Theory	100	100	100	100	400	
Practicals	50	50	50	50	200	

Course code	Practical Syllabus	Credits & lectures		
USMBP05	Based on USMB501 and USMB502 of Semester V	Credits 3 (8 periods/week) = 120 periods/semester		
USMBP06	Based on USMB503 and USMB504 of Semester V	Credits 3 (8 periods/week) = 120 periods/semester		

T.Y.B.Sc. Microbiology Practicals: Semester-V

T.Y.B.Sc. Microbiology Practicals: Semester-VI

Course code	Practical Syllabus	Credits & lectures		
USMBP07	Based on USMB601 and USMB602 of Semester VI	Credits 3 (8 periods/week) = 120 periods/semester		
USMBP08	Based on USMB603 and USMB604 of Semester VI	Credits 3 (8 periods/week) = 120 periods/semester		

COURSE WISE CREDIT ASSIGNMENT UNDER THE FACULTY OF SCIENCE

Program: B.Sc.

Course: Microbiology (USMB)

Course wise credit assignments under the faculty of science Type of Courses / Credits Assigned	First Year (Credit x No. of Courses)		Second Year (Credit x No. of Courses)		Third Year (Credit x No. of Courses)		Total
	First Semester	Second Semester	Third Semester	Fourth Semester	Fifth Semester	Sixth Semester	Credit Value
Core Courses (Theory)	04x03	04x03	06x02	06x02	2.5x04	2.5x04	68
Core Courses (Practicals)	02x03	02x03	03x02	03x02	1.5x04	1.5x04	36
Foundation course	02x01	02x01	02x01	02x01			08
Applied Component Courses (Theory)					02x01	02x01	04
Applied Component Courses (Practical)					02x01	02x01	04
Total	20	20	20	20	20	20	120