UNIVERSITY OF MUMBAI



Revised Syllabus for F.Y.B.Sc.
Program: B.Sc.
Course:MICROBIOLOGY(USMB)

(Choice Based Credit System with effect from the Academic year 2016-17)

PREAMBLE

With the introduction of Choice Based Credit System (CBCS) by the esteemed University from the academic year 2016-201, the existing syllabus of F.Y.B.Sc. Microbiology is restructured according to the CBCS pattern for its implementation from 2016-2017.

While earlier revision of the syllabus took care of balancing both the basic techniques and some of the advance techniques (as remaining will be introduced phase wise at S.Y.B.Sc. and T.Y.B.Sc level) in Microbiology, the present revision is related to restructuring of syllabus as per CBCS pattern.

The concepts of **Biosafety, Validation, Calibration and SOPs** have been introduced to make the learners aware about :-

- i. The biological hazards and safety measures
- ii. Importance of Validation and Calibration of Scientific equipments in industries and laboratories.
- iii. Writing of SOPs for instruments and their importance at work.

The unique chemistry of living systems results in large part from the remarkable and diverse properties of **Biomacromolecules**. Macromolecules from each of the four major classes may act individually in a specific cellular process, where as others associate with one another to form supramolecular structures. All of these structures are involved in important cellular processes. Since the arrival of information technology, biochemistry has evolved from an interdisciplinary role to becoming a core program for a new generation of interdisciplinary courses such as **bioinformatics and computational biochemistry**. Hence the module of macromolecules has been included in the revised syllabus to teach students the structure and function of biomolecules at an entry level with an objective to raise the student's awareness of the applicability of microcomputers in biochemistry as they go to the higher classes.

F.Y.B.Sc Microbiology Syllabus (General Outline) Revised for Choice Based Credit System To be implemented from the Academic year 2016-17

	SEMESTER I	
Course Code	Title	Credits
USMB-101	FUNDAMENTALS OF MICROBIOLOGY.	2 Credits
Theory		(45 lectures)
Unit-I	History, Introduction & Scope Of Microbiology Prokaryotic Cell Structure,	15 lectures.
Unit-II	Eukaryotic Cell Structure Biosafety In Microbiology	15 lectures.
Unit-III	Macromolecules	15 lectures.
USMB-102 Theory	BASIC TECHNIQUES IN MICROBIOLOGY.	2 Credits (45 lectures)
Unit-I	Microscopy & Staining	15 lectures.
Unit-II	Control Of Microorganisms	15 lectures.
Unit-III	Microbial Nutrition, Cultivation, Isolation & Preservation	15 lectures.
USMBP-1	PRACTICALS	2 Credits
	SECTION-1 FUNDAMENTALS OF MICROBIOLOGY. (Practicals Based On Unit-I,II & III Of USMB-101	1 Credit (45 lectures)
	SECTION-2 BASIC TECHNIQUES IN MICROBIOLOGY. (Practicals Based On Unit-I,II & III Of USMB-102	1 Credit (45 Lectures)
	SEMESTER II	
USMB-201 Theory	BASICS OF MICROBIOLOGY.	2 Credits (45 Lectures)
Unit-I	Study Of Different Groups Of Microbes-I	15 lectures.
Unit-II	Study Of Different Groups Of Microbes-II	15 lectures.
Unit-III	Microbial Growth	15 lectures.
USMB-202 Theory	EXPLORING MICROBIOLOGY.	2 Credits (45 Lectures)
Unit-I	Microbial Interactions	15 lectures.
Unit-II	Microbes & Human Health	15 lectures.
Unit-III	Advance Techniques In Microbiology & Instrumentation	15 lectures.
USMBP-2	PRACTICALS	2 Credits
	SECTION-1 BASICS OFMICROBIOLOGY. (Practicals Based On Unit-I,II & III Of USMB-201)	1 Credit (45 Lectures)
	SECTION-2 EXPLORING MICROBIOLOGY. (Practicals Based On Unit-I,II & III Of USMB-202)	1 Credit (45Lectures)

USMB-102 Theory	BASIC TECHNIQUES IN MICROBIOLOGY.	2 Credits (45 lectures)	Notional Periods Self Study (45)
Unit-I	Microscopy & Staining	15 lectures.	15
	1.1 Microscopy: History of microscopy, Optical spectrum, Lenses and mirrors: Simple and compound light microscope, Dark field Microscopy, Phase contrast	08 Lectures	
	 1.2 Staining procedures a. Dyes and stains: Types, Physicochemical basis Fixatives, Mordants, Decolorizers b. Simple and differential staining c. Special staining (Cell wall, Capsule, Lipid granules ,Spores, Metachromatic granules & Flagella) 	07 Lectures	
Unit-II	Control Of Microorganisms	15 lectures.	15
	2.1 Definition of frequently used terms & Rate of microbial death, Factors affecting the effectiveness of antimicrobial agents & Properties of an ideal disinfectant 2.2 Evaluation of disinfectant —Tube dilution & Agar plate techniques, Phenol coefficient, Tissue toxicity index 2.3 Physical methods of microbial control a. Dry & moist heat — mechanisms, instruments used and their operations b. Electromagnetic radiations — Ionizing radiations, mechanisms —advantages & disadvantages c. Bacteria proof filters d. Low temperature e. Osmotic pressure f. Desiccation 2.4 Chemical methods of microbial control — mechanism & advantages & disadvantages (if any) applications. a. Phenolics b. Alcohols c. Heavy metals and their compounds d. Halogens e. Quaternary ammonium compounds f. Halogens g. Dyes h. Surfaces active agents/Detergents i. Aldehydesp j. Peroxygens k. Sterilizing gases l.		

	2.5.Cl		
	2.5 Chemotherapeutic agents - List types of agents active		
	against various groups & mention the site of		
	action(Detailed mode of action not to be done)		
Unit-III	Microbial Nutrition, Cultivation, Isolation &	15 lectures.	15
	Preservation		
	3.1 Nutritional requirements – Carbon, Oxygen, Hydrogen,		
	Nitrogen, Phosphorus, Sulfur and growth factors.		
	3.2 Nutritional types of microorganisms		
	3.3 Types of Culture media with examples		
	* *		
	3.4 Isolation of microorganisms and pure culture		
	techniques		
	3.5 Preservation of microorganisms		
	3.6 Culture Collection Centres		
USMBP-1	PRACTICALS	2 Credits	Notional
			Periods
	SECTION-1	1 Credit	Self Study
	FUNDAMENTALS OF MICROBIOLOGY.	(45 lectures)	(45)
Unit-I	1. Assignment : Contribution of Scientists in the field		
Cint 1	of Microbiology		
	2. Special staining: Cell wall, capsule, endospore,		
TI.4 TT	flagella, lipid, metachromatic granules.		
Unit-II	3. Handling corrosive chemical using rubber teat		
	method for pipetting. Prevention of mouth pipetting		
	and use of auto-pipettes.		
	4. Discard of highly infectious pathogenic samples		
	like T.B, sputum etc.		
	5. Explain safety inoculation hood for infection		
	inoculations and laminar air flow.		
	6. On accidental spillage of/ breakage of culture		
	containers-precautions to be taken.		
	7. Demonstration of microbes in air, cough, on table		
	surface, finger tips.		
	8. Permanent slides of Eukaryotes & its organelles:		
	9. Assignment: Eukaryotic organelles		
Unit-III	10. Qualitative detection :		
Omt-111			
	11. Carbohydrates- Benedicts, Molisch's test.		
	12. Proteins, amino acids- Biuret, Ninhydrin.		
	13. Nucleic acid detection by DPA and Orcinol.	4.0 ***	0.150: 1
	SECTION-2	1 Credit	Self Study
	BASIC TECHNIQUES INMICROBIOLOGY.	(45 lectures)	(45)
Unit-I	1. Parts of a microscope,		
	2. Micrometry		
	3. Dark field and Phase contrast: Demonstration		
	4. Monochrome and differential staining procedures, Gram		
	staining& Negative Staining.		
Unit-II	5. Introduction to Laboratory equipments, disinfection &		
	discarding techniques in laboratory		
	6. Methods of preparation of glassware for Sterilization		
	10. Methods of preparation of glassware for stermzation	1	

USMB-202	EXPLORING MICROBIOLOGY.	2 Credits	Self Study
Theory		(45 lectures)	(45)
Unit-I	Microbial Interactions:	15 lectures.	15
	1.1 Types of Microbial Interactions : Mutulism,		
	Cooperation, Commensalisms, Predation Parasitism,		
	Amensalism, Competition		
	1.2 Human Microbe Interactions .		
	a) Normal flora of the human body: Skin, Nose &		
	Nasopharynx, Oropharynx, Respiratory tract, Eye,		
	External ear, Mouth, Stomach, Small intestine, Large		
	intestine, Genitourinary tract.		
	b) Relationship between microbiota& the host.		
	c) Gnotobiotic animals		
	1.3 Microbial associations with vascular plants		
	a) Phyllosphere		

	b) Rhizosphere & Rhizoplane		
	c) Mycorrhizae		
	d) Nitrogen fixation : Rhizobia, Actinorhizae,		
	Stem Nodulating Rhizobia		
	e) Fungal & Bacterial endophytes		
	f) Agrobacterium & other plant pathogens		
Unit-II	Microbes & Human Health:	15 lectures.	15
CIIIt-II	2.1 Difference between infection & disease.	13 lectures.	13
	Important terminology: Primary infection, secondary		
	infection.Contagious infection, occupational disorder,		
	clinical infection, subclinical infection, Zoonoses, genetic		
	disorder, vector borneinfection.		
	2.2 Factors affecting infection:		
	Microbial factors: adherence, invasion, role of virulence		
	factors in invasion, colonization & its effects.		
l	· ·		
	Host factors: natural resistance, species resistance, racial resistance.		
	2.3 Individual resistance : Factors influencing individual		
	resistance: Age, nutrition, personal hygiene, stress,		
	hormones, Addiction to drugs/ alcohol. Interaction between		
	Microbes & host is dynamic.		
	2.4 Host defense against infection: Overview		
	i) First line of Defence: for skin, respiratory tract,		
	gastrointestinal tract, genitourinary tract, eyes.		
	ii) Second line of defence: Biological barriers:		
	Phagocytosis, Inflammation		
	iii) Third line of defence: Brief introduction to antibody		
TI . 4 TTT	mediated & cell mediated immunity.	151 4	1.5
Unit-III	Advance Techniques In Microbiology & Instrumentation:	15 lectures.	15
	3.1 Electron Microscope:TEM,SEM,		
	3.2 Contrast enhancement for electron microscope		
	3.3 Fluorescent Microscope, Confocal Microscope		
	3.4 pH meter ,pH meter Validation and calibration		
	3.5 Colorimeter		
	3.6 Validation and calibration of Autoclave & Hot air Oven		
	3.7 Concepts :Laminar air flow systems, Biosafety cabinets		
	, Walk in Incubators, Industrial autoclaves, Cold Room.		
USMBP-2	PRACTICALS	2 Credits	
	SECTION-1	1 Credit	Self Study
	BASICS OFMICROBIOLOGY.	(45 lectures)	(45)
Unit-I	1. Spot assay and plaque assay of Bacteriophage		
	(Demonstration)		
	2. Slide Culture technique (Actinomycetes & Fungal		
	Culture)		
Unit-II	3.Isolation of yeast, cultivation of other fungi		

	4. Static & Shaker Cultures		
	5. Fungal Wet mounts & Study of Morphological		
	Characteristics: Mucor, Rhizopus, Aspergillus,		
	Penicillium,		
	6. Permanent slides of Algae, Protozoa		
Unit-III	7. Growth curve (Demonstration) only in complex media.		
	8.Breed's Count		
	9.Haemocytometer		
	10. Viable count: Spread plate and pour plate		
	11.Brown's opacity		
	12.Effect of pH and temperature on growth		
	13.Measurement of cell dimensions-Micrometry		
	SECTION-2	1 Credit	Self Study
	EXPLORING MICROBIOLOGY.	(45 lectures)	(45)
Unit-I	1. Normal flora of the Skin & Saliva		
	2. Wet Mount of Lichen		
	3. Bacteroid Staining & Isolation of Rhizobium		
	4. Azotobacter isolation & staining		
Unit-II	6.Study of virulence factors – Enzyme Coagulase		
	7.Study of virulence factors – Enzyme Hemolysin		
	8.Study of virulence factors – Enzyme Lecithinase		
Unit-III	9.Use of standard buffers for calibration and determination		
	of pH of a given solution		
	10.Determination of λ_{max} & Verification of Beer		
	Lambert's law		
	11.Determination & efficiency of Autoclave, Hot air oven		
	, LAF		
	12.Writing of SOP's for Instruments		
	13. Visit to a Microbiology laboratory in a research		
	Institute		

REFERENCES: USMB 101 & USMB 201

- 1. Prescott ,Hurley.Klein-Microbiology, 7th edition, International edition, McGraw Hill.
- 2. Kathleen Park Talaro& Arthur Talaro Foundations in Microbiology International edition 2002,| McGraw Hill.
- 3. Michael T.Madigan & J.M.Martin,Brock ,Biology of Microorganisms 12th Ed. Internationaledition 2006, Pearson Prentice Hall.
- 4. A.J.Salle, Fundamental Principles of Bacteriology.
- 5. Stanier.Ingraham et al ,General Microbiology 4th & 5th Ed. 1987, Macmillan Education Ltd
- 6. Microbiology TMH 5th Edition by Michael J.Pelczar Jr., E.C.S. Chan ,Noel R. Krieg
- 7. BIS:12035.1986: Code of Safety in Microbiological Laboratories

- 8. Outlines of Biochemistry 5/E, Conn P. Stumpf, G. Bruening and R. Doi. John Wiley & Sons. New York 1995
- 9. Lehninger. Principles of Biochemistry. 4th Edition. D. Nelson and M. Cox. W.H. Freeman and Company. New York 2005
- 10. Microbiology An Introduction. 6th Edition. Tortora, Funke and Case. Adisson Wesley Longman Inc. 1998.

REFERENCES: USMB 102& USMB 202

- 1. Microbiology TMH 5th Edition by Michael J.Pelczar Jr., E.C.S. Chan ,Noel R. Krieg
- 2. A.J.Salle, Fundamental Principles of Bacteriology, McGraw Hill Book Company Inc.1984
- 3. Cruikshank, Medical Microbiology, Vol -II
- 4. Prescott ,Hurley.Klein-Microbiology, 5th & 6th edition, International edition 2002 & 2006, McGraw Hill.
- 5. Michael T.Madigan & J.M.Martin,Brock ,Biology of Microorganisms 11th Ed. International edition ,2006, Pearson Prentice Hall.

MODALITY OF ASSESSMENT

Theory Examination Pattern:

(A) Semester End Theory Assessment -

100 Marks

- i. Duration These examinations shall be of **3 Hours** duration.
- ii. Theory question paper pattern:-
- 1. There shall be **four** questions. On each unit there will be one question with **25** Marks each & fourth one will be based on all the three units with **25** Marks.
- 2. All questions shall be **compulsory** with internal choice within the questions. Question 1 (Unit-I), Question 2 (Unit-II) & Question 3 (Unit-III) & Question 4 (combined units) will be of **50** Marks with internal options.
- 3. All Questions may be sub divided into sub questions of **five** marks objective questions and **twenty** marks of short or long questions of 5 to 10 marks each. Please ensure that the allocation of marks depends on the weightage of the topic

PRACTICAL EXAMINATION PATTERN

(B) External (Semester end practical examination): 50 Marks Per Section (Section-I based on course-1 & Section-II based on course-2)

Sr.No.	Particulars	Marks		Total
1.	Laboratory work (Section-I + Section-II)	40 + 40	=	80
2.	Journal	05 +05	=	10
3.	Viva	05 + 05	=	10

PRACTICAL BOOK/JOURNAL

Semester I:

The students are required to present a duly certified journal for appearing at the practical examination, failing which they will not be allowed to appear for the examination.

In case of loss of Journal and/ or Report, a Lost Certificate should be obtained from Head/ Co-ordinator / Incharge of the department; failing which the student will not be allowed to appear for the practical examination.

Semester II

The students are required to present a duly certified journal for appearing at the practical examination, failing which they will not be allowed to appear for the examination.

In case of loss of Journal and/ or Report, a Lost Certificate should be obtained from Head/ Co-ordinator / Incharge of the department; failing which the student will not be allowed to appear for the practical examination.

Overall Examination and Marks Distribution Pattern

Semester I

Course	USMB- 101	USMB- 102	Grand Total
Theory	100	100	200
Practicals	50	50	100

Semester II

Course	USMB- 201	USMB- 202	Grand Total
Theory	100	100	200
Practicals	50	50	100